REVIEW ARTICLE

Possible antithrombotic properties of propolis

Naoki Ohkura¹, Keiji Maruyama², Fumiko Kihara-Negishi³

¹Department of Medical and Pharmaceutical Sciences, School of Pharma-Sciences, Teikyo University, Tokyo, Japan ²Research Center for the Promotion of Pharmacy and Pharmaceutical Practice, School of Pharma-Sciences, Teikyo University, Tokyo, Japan ³Department of Life and Health Sciences, School of Pharma-Sciences, Teikyo University, Tokyo, Japan

ABSTRACT

The formation of thrombi in blood vessels results in thrombosis that is responsible for a considerable morbidity and mortality because it is associated with arterial diseases such as myocardial infarction, stroke, and peripheral occlusive disease in addition to venous thromboembolic disorders. Pharmaceutical agents such as anticoagulant agents and antiplatelet agents are applied for the prevention of the recurrence of thrombotic disorders. However, the use of these pharmaceuticals can result in side effects such as bleeding, as well as renal and hepatic disorders. The prevention and treatment of diseases using functional foods and alternative medicines have recently attracted an attention. Functional foods and alternative medicines with possible antithrombotic properties that have been used for many years are now receiving a significant focus in terms of the treatment and prevention of thrombosis. As they have already been used for so long, they are likely to be safe. Propolis is a hive product comprising resinous materials collected by bees from plants, and it includes various chemical compounds. Various biological activities of propolis have been indicated, and propolis is used as folk medicine and health supplement worldwide. This report reviews the possibility of using propolis as an antithrombotic agent.

Introduction

Blood normally flows through blood vessels unobstructed. Nevertheless, blood components such as cells and plasma can leak from wounds in damaged vessels, and severe bleeds can be fatal. Blood clots at wound sites of damaged vessels stop bleeding in a process called hemostasis that includes platelets and a cascade of coagulation factors [1]. In a broad sense, the fibrinolytic system and its component factors are a part of the hemostatic system because fibrinolysis removes clots that are no longer needed for hemostasis to complete [2]. Vascular endothelial cells that line the inside of blood vessels exert powerful antithrombotic actions that enable blood to circulate throughout the body [3]. However, blood clots also form at the sites of damaged vascular endothelium, where they can cause thrombus and

other thrombotic disorders. Such clots are called thrombi, and the situation involving them is called thrombosis [3]. Thrombi in the coronary arteries, lungs, and brain vessels can be fatal [4-6]. Namely, thrombosis is a pathological clot that results when hemostasis is excessively activated in the absence of bleeding. The clotting process in a healthy person is regulated as needed, and thrombus formation is avoided. Nevertheless, clots can form easily in the blood vessels of persons with risk factors such as dyslipidemia, diabetes mellitus, obesity, psychological stress, a sedentary lifestyle, and cigarette smoking [7–14]. These factors also increase the risk of atherosclerosis, in which blood vessels become clogged with fatty plaque deposits. Atherosclerosis additively increases the risk of thrombosis [15].

Contact Naoki Ohkura 🖾 n-ohkura@pharm.teikyo-u.ac.jp 🖬 Department of Medical and Pharmaceutical Sciences, School of Pharma-Sciences, Teikyo University, Tokyo, Japan.

© 2020 The Authors. This is an open access article under the terms of the Creative Commons Attribution NonCommercial ShareAlike 4.0 (https://creativecommons.org/licenses/by-nc-sa/4.0/).



Received August 07, 2019 Accepted January 05, 2020 Published February 22, 2020

KEYWORDS

Honeybee; Brazilian propolis; thrombosis; platelet aggregation; plasminogen activator inhibitor 1; caffeic acid phenethyl ester

∂ Open Access

The prevention of thrombosis has been prioritized in the developed countries, where (unhealthy) lifestyles have increased the risk of thrombotic events and the incidence of thrombosis [16,17]. Various drugs that are used to prevent thrombotic diseases in developed nations can prevent thrombosis [18], but these also prevent the hemostatic system from working properly, resulting in increased susceptibility to bleeding. Thus, milder agents with minimal side effects such as bleeding are desirable. The discovery of natural products and alternative medicines that have antithrombotic properties is now a significant target, and the application of such products to prevent thrombosis is anticipated [19–22]. In general, antithrombotic activity generally refers to antiplatelet action and the anticoagulant action of plasma and occasionally fibrinolytic activity [18]. Antithrombotic substances comprise anticoagulants that halt the coagulation system and interfere with further clot expansion, antiplatelet agents that decrease platelet aggregation and inhibit thrombus formation, and fibrinolytic enzymes that directly dissolve thrombus [23]. Whether or not these compounds exert antithrombotic effects through actions on blood coagulation factors, platelets and the fibrinolytic system should be experimentally assessed in vitro and in vivo. This review outlines the possible antithrombotic properties of propolis and natural substances found in propolis.

Chemical Composition and Representative Biological Activities

Chemical composition

Propolis is a hive product produced by mixing bee saliva with resinous materials that bees collect from various plants or confer trees [24]. Honeybee propolis is famous and popular; not only honeybees but also other types of bees produce propolis [25-30]. Propolis is a folk medicine and health supplement all over the world, and various biological activities have been indicated [24,31]. Propolis includes various chemical substances. The general ratios (%) of these substances are as follows: resins and balsam-50%, beeswax—30%, pollen—5%, essential and aromatic oils—10%, and other substances that include organic compounds [32]. In general, propolis contains polyphenols (flavonoids, phenolic acids, and esters), phenolic aldehydes, and ketones, among other compounds [32]. However, the composition and biological activities of propolis greatly depend on the location of the bees, the season when resins are collected. and the plant sources of the resins [33]. For example, propolis samples from Europe and North America mainly comprise flavonoids, phenolic acids, and their esters [34]. Brazilian propolis contains various biologically active organic compounds in abundance such as artepillin C [35], whereas Chinese propolis is highly antioxidative and abundant in benzyl caffeate [36]. Therefore, the chemical heterogeneity of propolis is easy to understand, and the effects of propolis are closely associated with the production area. Table 1 summarizes the geographic origins, main plant sources, and chemical compounds. Examples of seasonal differences are as follows: Isla et al. [37] reported the seasonal variations of the antioxidant activity of propolis from Argentina. Samples corrected in November had the most antioxidant activity. They found a correlation between antioxidant capacity and flavonoid content. Regueira et al. [38] showed different effects of the dry and rainy seasons on the antibacterial activity and chemical composition of Brazilian red propolis, and Souza et al. [39] found that seasonal variations influence the Mg, Fe, Na, Ca, and Cu contents of Brazilian propolis. Thus, the quality of propolis within the same region varies depending on the season.

Table 4	Coorena a la caria inco	manin plant any mana	مصاحبه ومتعاصية ومسامعته	
Table 1.	Geographic origins,	, main plant sources	and main bioactive c	ompounds.

Geographic origin	Plant source	Main bioactive compounds	Reference Nos.
Europe, North America, and temperate regions of Asia	<i>Populus</i> spp., most often <i>P. nigra</i> L.	Polyphenols, phenolic acids, and their esters	[34,87]
Russia	Betula verrucosa Ehrh.	Polyphenols	[87]
Brazil	<i>Baccharis</i> spp., predominantly <i>B.</i> <i>dracunculifolia</i> DC.	Prenylated cinnamic acid derivatives, Diterpenic acids	[35,88]
Cuba	Venezuela Clusia spp.	Polyprenylated benzophenones	[89]
Pacific region (Okinawa, Taiwan)	Unknown	C-prenylated flavanones, Furofuran lignans	[90]
Kenya	Unknown	Polyphenols	[91]
Greece and Cyprus	Unknown	Flavonoids, terpenes	[92]

Effects of propolis on the immune system

Propolis exerts biological activities, including effects on immune systems. Brazilian propolis increases tumor necrosis factor- α (TNF- α) production in mice [40]. Ethanol extracts of Brazilian propolis upregulate toll-like receptor 2 (TLR-2) and TLR-4 expression and interleukin-1 β (IL-1 β) production in Bagg Albino c (BALB/c) mice [41]. Sforcin et al. reported that the ethanol extract of Brazilian propolis enhances NK cell activity. A water extract of propolis induces IL-1 production by peritoneal macrophages *in vitro* [42], and propolis extracts stimulate neutrophil chemotactic activity [43,44].

Antioxidant activity

The wide spectrum of the biological activities of propolis largely results from its antioxidative effects. Many studies have investigated the antioxidant activities of propolis [45], and those of phenolic acids and flavonoids in propolis and relationships among their chemical structures have been reviewed [46]. The antioxidant activity of propolis is due to its abundant content of polyphenols such as phenolic acid and flavonoids [45]. The antioxidant activity of propolis greatly depends on the production area and the plant source from which it is derived. For example, the antioxidant activity of Brazilian red propolis is more potent than that of green and brown propolis [47–49]. Although most of the studies regarding the antioxidant properties of propolis have been in cell culture or experimental animals, a few studies have examined the antioxidant effects of propolis in humans [50–52].

Antimicrobial activity

Antimicrobial activity of propolis is very well documented. Propolis exerts antimicrobial activity against a wide range of bacteria, fungi, and viruses [24]. Propolis is more active against Gram-positive bacteria than Gram-negative bacteria [53,54]. It was shown that propolis from different geographic regions has a considerable antiviral activity by acting at different levels and interfering with the replication of some viruses such as herpes simplex types 1 and 2, adenovirus type 2, influenza virus, or human immunodeficiency virus [53]. Antifungal activity is also influenced by the chemical variation of propolis. Some studies have shown the effect of propolis from different geographic origins against different fungi [54].

Anti-cancer activity

Propolis is effective against cancers of the brain, head and neck, skin, breast, liver, pancreas, kidney,

bladder, prostate colon, and blood [55]. The key mechanism underlying the anticancer activity is thought to depend on the inhibition of matrix metalloproteinases, antiangiogenesis, metastasis prevention, cell cycle arrest, induction of apoptosis, and modulation of deleterious side effects induced by chemotherapy. The anticancer activities of propolis vary depending on the botanical source and geographic origin [56]. Brazilian red propolis possesses cytotoxic activities [57] against human hepatocellular carcinoma cell lines [58] and mouse skin tumors *in vivo* [59]. Greek propolis exerts antiproliferative activity against human colon adenocarcinoma cells (HT-29) [60].

Effects of propolis on oxidative stress

Oxidative stress is closely associated with diabetes, hypertension, and obesity [61-64], which might be the consequences and causes of the high rates of mortality due to cardiovascular diseases among humans and animals [65,66]. Propolis exerts inhibitory activities on oxidative stress. Yuan et al. showed the cytoprotective effects of Brazilian green propolis against oxidative stress induced by oxidized low-density lipoprotein in human umbilical vein endothelial cells (HUVECs) [67]. Sun et al. [68] reported the potential protective effects of bioactive constituents from Chinese propolis against acute oxidative stress induced by hydrogen peroxide in cardiac H9c2 cells. Mujica et al. [50] described the role of propolis in oxidative stress and lipid metabolism in a randomized controlled trial. Their study showed the effects of an orally administered propolis solution on the oxidative status and modulation of lipids in a human population [51]. These studies provide an important basis for the application of propolis to the prevention and treatment of cardiovascular diseases. These effects have been attributed to natural compounds in propolis [69].

Propolis polyphenol and its metabolism in the body

Polyphenols are classified according to their chemical structures as flavonoids, simple phenols, hydrolyzed tannins, and condensed tannins [70]. Flavonoids, simple phenols, and condensed tannins have attracted a notice as targets for functional food development, and propolis contains many flavonoids [71]. Many flavonoids exist in propolis as glycosides that comprise an aglycone combined with sugar, and the dynamics *in vivo* differ depending on the type of aglycone and its sugar chain. That is, when orally consumed polyphenols are taken up by intestinal epithelial cells, some glycosides are hydrolyzed by lactose-phlorizin hydrolase and P-glucosidase in intestinal epithelial cells to produce aglycone [72]. Thereafter, aglycones such as catechin and flavonol passively diffuse into the epithelium, and most of the aglycones taken into the intestinal epithelial cells that enter glucuronidation, sulfate conjugation, and methylation in the circulating blood. Although many points remain unclear about the mechanism(s) through which polyphenols exhibit various physiological functions and low tissue concentrations in vivo, polyphenols enter into the body through oral ingestion circulation in blood and exert various actions [72].

Antiplatelet Activity of Propolis

Platelets adhere to the disrupted surfaces of damaged blood vessels and release biologically active constituents that induce further aggregation to stop bleeding although platelets cannot self-aggregate under normal circumstances. Thus, platelets play important roles in not only hemostasis but also the pathogenesis of cardiovascular and cerebrovascular disorders [73]. Since activated platelets aggregate and form thrombus and platelet activation is relevant to various cardiovascular and cerebrovascular disorders, the inhibition of platelet activation is clinically important for patients with thrombosis, cardiovascular diseases, and cerebrovascular disorders. However, the antiplatelet agents that are currently applied to clinically treat and prevent these disorders are associated with many side effects, among which bleeding is the most critical. Safer antiplatelet drugs are needed for such situations. Medicines derived from natural products and alternative medicine such as propolis might offer solutions.

Caffeic acid phenethyl ester (CAPE) is an active component of propolis produced in Europe, the Far East, and New Zealand [71,74]. The findings of many studies *in vitro* and *in vivo* have shown that CAPE has diverse biological activities [74]. Caffeic acid phenethyl ester is thought to be responsible for the anticancer, antioxidant, immunomodulatory, antibacterial, antiviral, anti-inflammatory, neuroprotective, hepatoprotective, and cardioprotective effects of propolis [74,75]. Therefore, propolis might have a potential for various clinical applications.

Some studies have found that CAPE exerts the effects on platelet activation. Hsiao et al. [76] examined the influence of 15–100 pM CAPE in washed human platelets and platelet plug formation *in vivo* and found that its concentration dependently inhibits collagen-induced platelet activation.

Platelet aggregation stimulated by the glycoprotein VI agonist, convulxin, as well as the $\alpha_{\alpha}\beta_{\alpha}$ integrin agonist, aggretin, is also inhibited by CAPE (25 µM). The inhibition of collagen-induced platelet activation by CAPE is accompanied by [Ca²⁺] mobilization, phosphoinositide breakdown, the activation of protein kinase C, and mitogen-activated protein kinases such as extracellular signal-regulated kinase 2 (ERK 2), c-Jun amino-terminal kinase, and p38 mitogen-activated protein kinase (MAPK), as well as v-Akt murine thymoma viral oncogene (AKT) phosphorylation, and thromboxane A (TXA₂) formation. In addition, CAPE (25 µM) interferes with FITC-collagen binding to platelet membranes. Platelet inhibition by CAPE is, at least in part, mediated by binding to collagen receptors such as a2pi integrin and GP VI [76]. Zhou et al. [77] showed that a CAPE analog (CAPE-NO₂) inhibits collagen-induced platelet aggregation, and they also suggested that this is associated with the downregulation of thromboxane B₂ (TXB₂), cyclooxygenase 1 (COX-1), and 5-hydroxytryptamine (SHT) and the elevation of NO and cyclic guanosine monophosphate (cGMP) [77]. Zhang et al. [78] found that an aqueous extract of propolis dose dependently inhibited the platelet aggregation induced by the agonists, adenosine diphosphate (ADP), thrombin receptor activator peptide, and collagen. Among CAPE, galangin, apigenin, quercetin, kaempferol, ferulic acid, rutin, chrysin, pinostrobin, and pinocembrin are the components of propolis; only CAPE, galangin, and pinostrobin inhibited platelet aggregation. These findings indicated that propolis components including CAPE might have therapeutic value for fighting thrombotic disease. One study in vivo showed that CAPE (5 mg/kg) significantly prolonged the latency of platelet plug induction in mice [76], but clinical findings have not been reported.

Bojic et al. recently showed that the ethanolic extracts of propolis reduce ADP-induced platelet aggregation using whole-blood platelet aggregation assays [79]. Their study showed the antiaggregatory potential of propolis ethanolic extracts in low micromolar concentrations on whole blood samples. Martina et al. [80] found that oral propolis (65 mg/kg/day) prolonged tail bleeding time using mouse tail bleeding assays, which reflects platelet aggregation activity *in vivo*. They showed that the activity of oral aspirin (10.4 mg/kg) was similar. Although further studies are needed to confirm the beneficial effects *in vivo*, these findings suggested that propolis supplementation can influence platelet aggregation and consequently thrombus formation and might have the potential to prevent cardiovascular diseases if proven in human studies.

Effects of Propolis on Fibrinolytic System

Fibrinolysis is a process in which clot degradation is modulated after damaged vascular tissue is repaired and replaced. The fibrinolytic system removes fibrin from the vascular system and thus prevents the enlargement of pathogenic hemostatic clots and vessel occlusion [2]. Therefore, attenuating fibrinolysis can lead to an increased risk of thrombosis. Fibrinolysis is regulated by plasminogen activator (PA) and plasminogen activator inhibitor 1 (PAI-1) [81]. The liver, adipose tissues, muscle, bone, and hematopoietic cells produce PAI-1, which inhibits tissue plasminogen activator during blood fibrinolysis [82]. Thrombus becomes difficult to dissolve when plasma PAI-1 concentrations are elevated, and persistent blood clots lead to thrombosis. Since plasma PAI-1 is elevated in patients with metabolic syndrome including obesity and diabetes, PAI-1 might be associated with a thrombotic tendency in such patients [82]. That is, a link between PAI-1 and metabolic syndrome has been established, and elevated plasma PAI-1 levels are now considered as a true component of the syndrome.

Chronic low-grade inflammation has been linked to the progression of obesity and related diseases [83]. Elevated plasma PAI-1 is closely associated with chronic inflammation in the adipose tissues of obese patients. Therefore, controlling PAI-1 elevation associated with chronic low-grade inflammation might help to prevent thrombosis caused by lifestyle-related diseases. Low-grade inflammation was used to induce a thrombotic tendency in an animal model and found that an orally administered ethanol extract of Brazilian propolis inhibits plasma PAI-1 elevation during inflammation [84]. The ethanol extract of Brazilian propolis suppressed PAI-1 production in cultured HUVEC stimulated with inflammatory TNF- α that increases PAI-1 release into the medium [85]. Chrysin suppresses the TNFαinduced increase in PAI-1 secretion, and chrysin is the most potent inhibitor of Brazilian propolis in terms of PAI-1 release from HUVEC [86]. However, as the content of chrysin in Brazilian propolis is very low, other contents of orally consumed Brazilian propolis might also be associated with the suppression of PAI-1 production. Further studies are needed to elucidate how propolis affects PAI-1 production and to identify specific molecules in Brazilian propolis that suppresses an increase in PAI-1.

Application of Propolis to the Prevention and Therapy of Thrombosis

Although studies of the effects of propolis on thrombosis are relatively rare, expectations that propolis will be able to treat and prevent thrombosis are enhanced. We considered that the possible antithrombotic properties of propolis should be announced to other investigators to further develop this field.

As described above, antithrombotic actions are progressed on suppressing platelet aggregation and PAI-1 production. Table 2 summarizes these antithrombotic effects of propolis. Studies using whole blood have shown that adding a small amount

Table 2. Antithrombo	tic effects of	propolis and	compounds in propoli	s.

Effects	Experimental target	Geographic origin of propolis	Active compounds	Reference Nos.
Inhibition of platelet aggregation	Isolated platelets	China	Unknown (water extract)	[78]
Inhibition of platelet aggregation	Isolated platelets	-	CAPE	[76]
Inhibition of platelet aggregation	Isolated platelets	-	CAPE analog	[77]
Inhibition of platelet aggregation	Platelet in whole blood	Various regions	Unknown (ethanol extract)	[79]
Prolonged tail bleeding	Mice	Unknown	Unknown	[80]
Inhibition of PAI-1 production from endothelial cells	Cultured endothelial cells	Brazil	Chrysin	[85,86]
Attenuation of PAI-1 production	Mice	Brazil	Unknown	[84]

of propolis (5 μ g in terms of flavonoids) to whole blood inhibits platelet aggregation [79]. Oral propolis (65 mg/kg/day) prolongs tail bleeding time that reflects platelet aggregation activity *in vivo* [80]. In addition, food containing 0.5% ethanol extract of propolis inhibits PAI-1 production in mice [84]. To date, the antithrombotic effects of propolis have not been clinically studied, but the propolis concentrations used in these reports will be useful for future clinical studies.

Similar to other biological properties, the antithrombotic properties of propolis should be directly associated with its chemical configuration, which varies according to regional vegetation, pollen collection season, collection techniques, and bee species. Active components in propolis and their effects on blood coagulation factors, platelets, and fibrinolytic system will require further investigation before propolis can be clinically applied. The validation and quality control of propolis that possesses antithrombotic properties are also important for clinical applications.

References

- [1] Rasche H. Haemostasis and thrombosis: an overview. Eur Heart J Suppl 2001; 3(suppl_Q):Q3–7.
- [2] Chapin JC, Hajjar KA. Fibrinolysis and the control of blood coagulation. Blood Rev 2015; 29(1):17–24.
- [3] Wang M, Hao H, Leeper NJ, Zhu L. Early Career Committee. Thrombotic regulation from the endothelial cell perspectives. Arterioscler Thromb Vasc Biol 2018; 38(6):e90–5.
- [4] Lacey MJ, Raza S, Rehman H, Puri R, Bhatt DL, Kalra A. Coronary embolism: a systematic review. Cardiovasc Revasc Med 2019; pii; S1553-8389(19)30311-2.
- [5] Yang C, Zhu L. Sudden death caused by acute pulmonary embolism after laparoscopic total extraperitoneal inguinal hernia repair: a case report and literature review. Hernia 2017; 21(3):481–6.
- [6] Virmani R, Burke AP, Farb A. Sudden cardiac death. Cardiovasc Pathol 2001; 10(5):211–8.
- [7] Pechlivani N, Ajjan RA. Thrombosis and vascular inflammation in diabetes: mechanisms and potential therapeutic targets. Front Cardiovasc Med 2018; 19(5):1.
- [8] Gaiz A, Mosawy S, Colson N, Singh I. Thrombotic and cardiovascular risks in type two diabetes; Role of platelet hyperactivity. Biomed Pharmacother 2017; 94:679–86.
- [9] Henning RJ. Type-2 diabetes mellitus and cardiovascular disease. Future Cardiol 2018; 14(6):491–509.
- [10] Austin AW, Wissmann T, von Kanel R. Stress and hemostasis: an update. Semin Thromb Hemost 2013; 39(8):902–12.

- [11] Bairey Merz CN, Dwyer J, Nordstrom CK, Walton KG, Salerno JW, Schneider RH. Psychosocial stress and cardiovascular disease: pathophysiological links. Behav Med 2002; 27(4):141–7.
- [12] Cannegieter SC. Travel-related thrombosis. Best Pract Res Clin Haematol 2012; 25(3):345–50.
- [13] Gavish I, Brenner B. Air travel and the risk of thromboembolism. Intern Emerg Med 2011; 6(2):113–6.
- [14] Gurbel PA, Baker BA, Bailey WL, Bliden KP, Tantry US. Unravelling the smokers' paradox: cigarette smoking, high-risk coronary artery disease and enhanced clinical efficacy of oral P2Y12 inhibitors. Thromb Haemost 2014; 111(6):1187–90.
- [15] Al Said S, Bode C, Duerschmied D. Anticoagulation in atherosclerotic disease. Hamostaseologie 2018; 38(4):240–6.
- [16] Pikovsky O, Rabinovich A. Prevention and treatment of the post-thrombotic syndrome. Thromb Res 2018; 164:116–24.
- [17] Katsiki N, Purrello F, Tsioufis C, Mikhailidis DP. Cardiovascular disease prevention strategies for type 2 diabetes mellitus. Expert Opin Pharmacother 2017; 18(12):1243–60.
- [18] Antithrombotic Agents. LiverTox: clinical and research information on drug-induced liver injury [Internet]. National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda MD, 2012–2018.
- [19] Mousa SA. Antithrombotic effects of naturally derived products on coagulation and platelet function. Methods Mol Biol 2010; 663:229–40.
- [20] McEwen BJ. The Influence of Herbal Medicine on Platelet Function and Coagulation. A narrative review. Semin Thromb Hemost 2015; 41:300–14.
- [21] Yamamoto J, Yamada K, Naemura A, Yamashita T, Arai R. Testing various herbs for antithrombotic effect. Nutrition 2005; 21:580–7.
- [22] Chen C, Yang FQ, Zhang Q, Wang FQ, Hu YJ, Xia ZN. Natural products for antithrombosis. Evid Based Complement Alternat Med 2015; 2015:876426.
- [23] Tsoucalas G, Chevallier J, Karamanou M, Papaioannou T, Sgantzos M, Androutsos G. Historical hallmarks of anticoagulation and antiplatelet agents. Curr Pharm Des 2016; 22(13):1857–61.
- [24] Khalil ML. Biological activity of bee propolis in health and disease. Asian Pac J Cancer Prev 2006; 7:22–31.
- [25] Torres AR, Sandjo LP, Friedemann MT, Tomazzoli MM, Maraschin M, Mello CF, Santos ARS. Chemical characterization, antioxidant and antimicrobial activity of propolis obtained from Melipona quadrifasciata quadrifasciata and Tetragonisca angustula stingless bees. Braz J Med Biol Res 2018; 51(6):e7118.
- [26] Santos HFD, Campos JF, Santos CMD, Balestieri JBP, Silva DB, Carollo CA, et al. Chemical profile and antioxidant, anti-inflammatory, antimutagenic and antimicrobial activities of geopropolis from the

stingless bee Melipona orbignyi. Int J Mol Sci 2017; 18(5). pii: E953.

- [27] Bonamigo T, Campos JF, Alfredo TM, Balestieri JB, Cardoso CA, Paredes-Gamero EJ, et al. Antioxidant, cytotoxic, and toxic activities of propolis from two native bees in Brazil: Scaptotrigona depilis and Melipona quadrifasciata anthidioides. Oxid Med Cell Longev 2017; 2017:1038153.
- [28] Kustiawan PM, Phuwapraisirisan P, Puthong S, Palaga T, Arung ET, Chanchao C. Propolis from the Stingless Bee Trigona incisa from East Kalimantan, Indonesia, induces in vitro cytotoxicity and apoptosis in cancer cell lines. Asian Pac J Cancer Prev 2015; 16(15):6581–9.
- [29] Campos JF, Dos Santos UP, da Rocha Pdos S, Damiao MJ, Balestieri JB, Cardoso CA, et al. Antimicrobial, antioxidant, anti-inflammatory, and cytotoxic activities of propolis from the stingless bee Tetragonisca fiebrigi (Jatai). Evid Based Complement Alternat Med 2015; 2015:296186.
- [30] Dutra RP, Abreu BV, Cunha MS, Batista MC, Torres LM, Nascimento FR, et al. Phenolic acids, hydrolyzable tannins, and antioxidant activity of geopropolis from the stingless bee Melipona fasciculata Smith. J Agric Food Chem 2014; 62(12):2549–57.
- [31] Banskota AH, Tezuka Y, Kadota S. Recent progress in pharmacological research of propolis. Phytother Res 2001; 15:561–71.
- [32] Anjum SI, Ullah A, Khan KA, Attaullah M, Khan H, Ali H, et al. Composition and functional properties of propolis (bee glue): a review. Saudi J Biol Sci 2019; 26(7):1695–703.
- [33] Bankova V. Chemical diversity of propolis and the problem of standardization. J Ethnopharmacol 2005; 100:114–7.
- [34] Burdock GA. Review of the biological properties and toxicity of bee propolis (propolis). Food Chem Toxicol 1998; 36:347–63.
- [35] Salatino A, Teixeira EW, Negri G, Message D. Origin and chemical variation of Brazilian propolis. Evid Based Complement Alternat Med 2005; 2:33–8.
- [36] Yamauchi R, Kato K, Oida S, Kanaeda J, Ueno Y. Benzyl caffeate, an antioxidative compound isolated from propolis. Biosci Biotechnol Biochem 1992; 56:1321–2.
- [37] Isla MI, Zampini IC, Ordonez RM, Cuello S, Juarez BC, Sayago JE, et al. Effect of seasonal variations and collection form on antioxidant activity of propolis from San Juan, Argentina. J Med Food 2009; 12(6):1334–42.
- [38] Regueira MS Neto, Tintino SR, da Silva ARP, Costa MDS, Boligon AA, Matias EFF, et al. Seasonal variation of Brazilian red propolis: Antibacterial activity, synergistic effect and phytochemical screening. Food Chem Toxicol 2017; 107(Pt B):572–80.
- [39] Souza EA, Zaluski R, Veiga N, Orsi RO. Effects of seasonal variations and collection methods on the mineral composition of propolis from Apis

mellifera Linnaeus Beehives. Braz J Biol 2016; 76(2):396–401.

- [40] da Silva SS, Thome Gda S, Cataneo AHD, Miranda MM, Felipe I, Andrade CGTJ, et al., Brazilian propolis antileishmanial and immunomodulatory effects. Evid Based Complement Altern Med 2013; 2013:673058.
- [41] Orsatti CL, Missima F, Pagliarone AC, Sforcin JM, Th1/Th2 cytokines' expression and production by propolis-treated mice. J Ethnopharmacol 2010; 129:314–8.
- [42] Dimov V, Ivanovska N, Bankova V, Popov S. Immunomodulatory action of propolis: IV. Prophylactic activity against gram-negative infections and adjuvant effect of the water-soluble derivative. Vaccine 1992; 10:817–23.
- [43] Quiroga EN, Sampietro DA, Soberon, JR, Sgariglia MA, Vattuone MA. Propolis from the northwest of Argentina as a source of antifungal principles. J Appl Microbiol 2006; 101:103–10.
- [44] Sampietro DA, Sampietro Vattuone MM, Vattuone MA. Immunomodulatory activity of Apis mellifera propolis from the North of Argentina. LWT—Food Sci Technol 2016; 70:9–15.
- [45] Kocot J, Kielczykowska M, Luchowska-Kocot D, Kurzepa J, Musik I. Antioxidant potential of propolis, bee pollen, and royal jelly: possible medical application. Oxid Med Cell Longev 2018; 2018:7074209.
- [46] Kurek-Gorecka A, Rzepecka-Stojko A, Gorecki M, Stojko J, Sosada M, Swierczek-Zieba G. Structure and antioxidant activity of polyphenols derived from propolis. Molecules 2013; 19:78–101.
- [47] Alencar SM, Oldoni TLC, Castro ML, Cabral ISR, Costa-Neto CM, Cury JA, et al. Chemical composition and biological activity of a new type of Brazilian propolis: red propolis. Ethnopharmacol 2007; 113:278–83.
- [48] de Mendonça IC, Porto IC, do Nascimento TG, de Souza NS, Oliveira JM, Arruda RE, et al. Brazilian red propolis: phytochemical screening, antioxidant activity and effect against cancer cells. BMC Complement Altern Med 2015; 15:357.
- [49] Machado BA, Silva RP, Barreto Gde A, Costa SS, Silva DF, Brandao HN, et al. Chemical composition and biological activity of extracts obtained by supercritical extraction and ethanolic extraction of brown, green and red propolis derived from different geographic regions in Brazil. PLoS One 2016; 11:e0145954.
- [50] Mujica V, Orrego R, Perez J, Romero P, Ovalle P, Zuniga-Hernandez J, et al. The role of propolis in oxidative stress and lipid metabolism: a randomized controlled trial. Evid Based Complement Alternat Med. 2017; 2017:4272940.
- [51] Jasprica I, Mornar A, Debeljak Z, Smolcic-Bubalo A, Medic-Saric M, Mayer L, et al. In vivo study of propolis supplementation effects on antioxidative

status and red blood cells. J Ethnopharmacol 2007; 110(3):548–54.

- [52] Zhao L, Pu L, Wei J, Li J, Wu J, Xin Z, Gao W, Guo C. Brazilian green propolis improves antioxidant function in patients with type 2 diabetes mellitus. Int J Environ Res Public Health 2016; 13(5). pii: E498.
- [53] Fokt H, Pereira A, Ferreira AM, Cunha A, Aguiar C. "How do bees prevent hive infections? The antimicrobial properties of propolis. In: Mendez-Vilas A (ed.). Current research, technology and education topics in applied microbiology and microbial biotechnology, vol. 1, Microbiology Book Series Number 2, pp 481–93, 2010.
- [54] Kujumgiev A, Tsvetkova I, Serkedjieva Y, Bankova V, Christov R, Popov S. Antibacterial, antifungal and antiviral activity of propolis of different geographic origin. J Ethnopharmacol 1999; 64(3):235–40.
- [55] Premratanachai P, Chanchao C. Review of the anticancer activities of bee products. Review. Asian Pac J Trop Biomed 2014; 4(5):337–44.
- [56] Patel S. Emerging adjuvant therapy for cancer: propolis and its constituents. J Diet Suppl 2016; 13:245–68.
- [57] Li F, Awale S, Tezuka Y, Kadota S. Cytotoxic constituents from Brazilian red propolis and their structure- activity relationship. Bioorg Med Chem 2008; 16:5434–40.
- [58] Matsuno T, Saito M, Matsumoto Y, Morikawa J. A new benzo-gamma-pyran derivative isolated from propolis. Z Naturforsch 1998; 53:1037–39.
- [59] Mitamura T, Matsuno T, Sakamoto S, Maemura M, Kudo H, Suzuki S, et al. Effects of a new clerodane diterpenoid isolated from propolis on chemically induced skin tumors in mice. Anticancer Res 1995; 16:2669–72.
- [60] Pratsinis H, Kletsas D, Melliou E, Chinou I. Antiproliferative activity of Greek propolis. J Med Food 2010; 13:286–90.
- [61] Shen GX. Lipid disorders in diabetes mellitus and current management. Curr Pharm Anal 2007; 3:17–24.
- [62] Orsolic N, Basic I. Honeybee products and their polyphenolic compounds in treatment of diabetes. In Phytopharmacology and Therapetutic Values IV. Govil JN, Singh VK Eds., Studium Press LLC, Houston, TX, Volume 22, pp. 455–553, 2008.
- [63] Orsolic N, Sirovina D, Koncic MZ, Lackovic, G.; Gregorovic, G. Effect of Croatian propolis on diabetic nephropathy and liver toxicity in mice. BMC Complement Altern Med 2012; 12:117.
- [64] Brzovic-Saric V, Landeka I, Saric B, Barberic M, Andrijasevic L, Cerovski B, et al. Levels of selected oxidative stress markers in the vitreous and serum of diabetic retinopathy patients. Mol Vis 2015; 21:649–64.

- [65] Orsolic N, Kunstic M, Kukolj M, Gracan R, Nemrav J. Oxidative stress, polarization of macrophages and tumour angiogenesis: Efficacy of caffeic acid. Chem Biol Interact 2016; 256:111–24.
- [66] Althunibat, OY, Al Hroob, AM, Abukhalil MH, Germoush MO, Bin-Jumah M, Mahmoud AM. Fisetin ameliorates oxidative stress, inflammation and apoptosis in diabetic cardiomyopathy. Life Sci 2019; 221:83–92.
- [67] Yuan W, Chang H, Liu X, Wang S, Liu H, Xuan H. Brazilian green propolis inhibits Ox-LDL—stimulated oxidative stress in human umbilical vein endothelial cells partly through PI3K/Akt/ mTOR—mediated Nrf2/HO-1 pathway. Evid Based Complement Alternat Med 2019; 2019:5789574.
- [68] Sun L, Wang K, Xu X, Ge M, Chen Y, Hu F. Potential protective effects of bioactive constituents from Chinese propolis against acute oxidative stress induced by hydrogen peroxide in cardiac H9c2 cells. Evid Based Complement Alternat Med 2017; 2017:7074147.
- [69] Mehta J, Rayalam S, Wang X. Cytoprotective effects of natural compounds against oxidative stress. Antioxidants (Basel) 2018; 7(10). pii: E147.
- [70] Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci 2016; 5:e47.
- [71] Huang S, Zhang CP, Wang K, Li GQ, Hu FL. Recent advances in the chemical composition of propolis. Molecules 2014; 19(12):19610–32. Review.
- [72] Serreli G, Deiana M. In vivo formed metabolites of polyphenols and their biological efficacy. Food Funct 2019; 10(11):6999–7021.
- [73] Badimon L, Vilahur G. Platelets, arterial thrombosis and cerebral ischemia. Cerebrovasc Dis 2007; 24 Suppl 1:30–9.
- [74] Tolba MF, Azab SS, Khalifa AE, Abdel-Rahman SZ, Abdel-Naim AB. Caffeic acid phenethyl ester, a promising component of propolis with a plethora of biological activities: a review on its antiinflammatory, neuroprotective, hepatoprotective, and cardioprotective effects. IUBMB Life 2013; 65:699–709.
- [75] Murtaza G, Karim S, Akram MR, Khan SA, Azhar S, Mumtaz A, Bin Asad MH. Caffeic acid phenethyl ester and therapeutic potentials. Biomed Res Int 2014; 2014:145342.
- [76] Hsiao G, Lee JJ, Lin KH, Shen CH, Fong TH, Chou DS, Sheu JR. Characterization of a novel and potent collagen antagonist, caffeic acid phenethyl ester, in human platelets: *in vitro* and *in vivo* studies. Cardiovasc Res 2007; 75:782–92.
- [77] Zhou K, Li X, Du Q, Li D, Hu M, Yang X, Jiang Q, Li Z. A CAPE analogue as novel antiplatelet agent efficiently inhibits collagen-induced platelet aggregation. Pharmazie 2014; 69:615–20.
- [78] Zhang YX, Yang TT, Xia L, Zhang WF, Wang JF, Wu YP. Inhibitory effect of propolis on platelet aggregation in vitro. J Healthc Eng 2017; 2017:3050895.

- [79] Bojic M, Antolic A, Tomicic M, Debeljak Z, Males Z. Propolis ethanolic extracts reduce adenosine diphosphate induced platelet aggregation determined on whole blood. Nutr J 2018; 17(1):52.
- [80] Martina SJ, Luthfi M, Govindan P, Wahyuni AS. Effectivity comparison between aspirin, propolis, and bee pollen as an antiplatelet based on bleeding time taken on mice. MATEC Web of Conferences 197, 07008 (2018):1–7.
- [81] Alessi MC, Nicaud V, Scroyen I, Lange C, Saut N, Fumeron F, et al. Association of vitronectin and plasminogen activator inhibitor-1 levels with the risk of metabolic syndrome and type 2 diabetes mellitus. Results from the D.E.S.I.R. prospective cohort. Thromb Haemost 2011; 106:416–22.
- [82] Van De Craen, Declerck PJ, Gils A. The biochemistry, physiology and pathological roles of PAI-1 and the requirements for PAI-1 inhibition in vivo. Thromb Res 2012; 130:576–85.
- [83] Longo M, Zatterale F, Naderi J, Parrillo L, Formisano P, Raciti GA, Beguinot F, Miele C. Adipose tissue dysfunction as determinant of obesity-associated metabolic complications. Int J Mol Sci 2019:13;20(9). pii: E2358.
- [84] Ohkura N, Oishi K, Kihara-Negishi F, Atsumi G, Tatefuji T Effects of a diet containing Brazilian propolis on lipopolysaccharide induced increases in plasma plasminogen activator inhibitor-1 levels in mice. J Intercult Ethnopharmacol 2016; 5:439–43.
- [85] Ohkura N, Takata Y, Ando K, Kanai S, Watanabe E, Atsumi G Propolis and one of its constituent chrysin inhibit plasminogen activator inhibitor-1

production induced by tumor necrosis factor-alpha and lipopolysaccharide. J Apicult Res 2012; 51:179–84.

- [86] Ohkura N, Ando K, Takata Y, Kanai S, Ishibashi K, Taniguch M, Tatefuji T Atsumi GI Positions of hydroxyl groups in chrysin are critical for inhibiting plasminogen activator inhibitor-1 release from human umbilical vein endothelial cells. Nat Prod Commun 2017; 12:499–502.
- [87] Monti M, Berti E, Carminati G, Cusini M. Occupational and cosmetic dermatitis from propolis. Contact Dermatitis 1983; 9(2):163.
- [88] Marcucci M. Propolis: chemical composition, biological properties and therapeutic activity. Apidologie 1995; 26(2):83–99.
- [89] Trusheva B, Popova M, Bankova V, Tsvetkova I, Naydensky C, Sabatini AG. A new type of European propolis containing bioactive labdanes. Rivista Italiana EPPOS 2003; 36:3–7.
- [90] Kumazawa S, Hayashi K, Kajiya K, Ishii T, Hamasaka T, Nakayama T. Studies of the constituents of Uruguayan propolis. J Agric Food Chem 2002; 50(17):4777–82.
- [91] Kosalec I, Pepeljnjak S, Bakmaz M, Vladimir-Knezevic S. Flavonoid analysis and antimicrobial activity of commercially available propolis products. Acta Pharmaceutica 2005; 55(4):423–30.
- [92] Kalogeropoulos N, Konteles SJ, Troullidou E, Mourtzinos I, Karathanos VT. Chemical composition, antioxidant activity and antimicrobial properties of propolis extracts from Greece and Cyprus. Food Chem 2009; 116(2):452–61.